

# RIDA<sup>®</sup>TUBE

Art. No. GZ3013



R-Biopharm AG, An der neuen Bergstraße 17, D-64297 Darmstadt, Germany

Tel.: +49 (0) 61 51 81 02-0 / Telefax: +49 (0) 61 51 81 02-20



## 1. Intended use

For *in-vitro* diagnostics. Stool collection tubes (without buffer) for the stool sample taking and extraction in laboratories.

## 2. Kit content

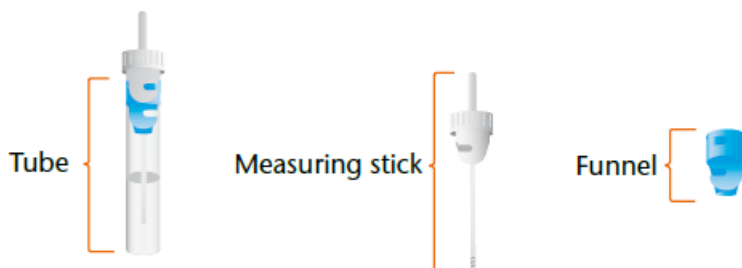
One kit contains 50 stool sampling tubes.

## 3. Description of stool sampling tube

One RIDA<sup>®</sup>TUBE is composed of the following parts:

- tube
- measuring stick with measuring tip
- funnel

### Setup:



## 4. Storage

The stool collection tubes need to be stored at room temperature.

## 5. Necessary equipment

Vortex, pipettes, inoculation loop or wooden stick

## **6. Warning/ precautions**

We do not recommend centrifuging the tube.

Stool samples should be treated as potentially infectious material.

## **7. Collection and storage of samples**

The collection and storage of the samples is determined by the type of analyte, its stability and the used extraction buffer.

With the measuring tip 10 mg stool can be taken. The tube can take a maximal volume of 2.5 ml.

## **8. Sample preparations**

The stool sample should be at room temperature (20 – 25 °C) before extraction and should be homogenized before sample taking e.g. with an inoculation loop or a wooden stick.

When transferred to the stool collection tube the grooves of the measuring tip have to be completely filled with stool. Moreover, no stool should be at the bar of the measuring stick.

Before test implementation, the tube has to be vortexed until the entire stool, taken with the measuring tip is suspended in extraction buffer.

## 9. Collecting samples with sample collection device – instruction for use

### 9.1 Filling of the RIDA<sup>®</sup>TUBE with buffer

Please notice: With the measuring tip 10 mg stool sample can be taken. The tube can take a maximal volume of 2.5 ml.

1. Pre-filling of the stool collection tubes before stool sampling is recommended.
2. Therefor open the tube at the blue funnel by turning it counterclockwise. Remove the blue funnel, as well as the white measuring stick from the tube.
3. After filling the tubes with the requested buffer, the tube has to be closed with the blue funnel as well as the measuring stick. The storage conditions of the filled tube depend on the storage conditions of the used buffer.

### 9.2 Sample taking of the RIDA<sup>®</sup>TUBE with buffer

1. Remove the white measuring stick from the tube by turning it counterclockwise. When you open the tube grasp the blue stopper between your thumb and forefinger. The blue stopper has to remain at the RIDA<sup>®</sup>TUBE.
2. Dip the measuring tip into the stool specimen at three different places.
3. Make sure that the grooves of the measuring tip are filled with stool.
4. Return the stick into the tube. By inserting the stick into the tube, excess stool material remains in the blue funnel insert. Close the tube carefully by turning the white measuring stick clockwise. In case of liquid stool samples, 10 µl can be pipetted into the stool collection tube.
5. Shake the tube by vortexing prior to testing. The stool material has to be suspended completely in extraction buffer. In case the stool is very hard it is recommended to tip the tube softly onto a hard surface until the sample is fully removed from the measuring tip.
6. Let the stool extract sediment. The RIDA<sup>®</sup>TUBE shouldn't be centrifuged.
7. For testing, open the tube at the blue shutter and remove it together with the measuring stick from the tube. The stool extract can now be used for further testings.

Note: The customer shall be responsible for the validation of the RIDA<sup>®</sup>TUBE for use with further IVD-products. The RIDA<sup>®</sup>TUBE can also be used on automatic 4-Plate-ELISA-Systems, e.g. DSX or DS2 from Dynex. Please contact R-Biopharm AG or your local distributor, if you are interested in using the RIDA<sup>®</sup>TUBE on automatic 4-Plate-ELISA-Systems.

## **10. Storage of extracts**

The storage of the extract depends on the examined analyte as well as its stability in the used extraction buffer.